

## Two New Alkaloids from the Bark of *Sarcomelicope megistophylla*

Magdalene Papageorgiou,<sup>†</sup> Nikolas Fokialakis,<sup>†</sup> Sofia Mitaku,<sup>†</sup> Alexios-Leandros Skaltsounis,<sup>\*,†</sup> François Tillequin,<sup>‡</sup> and Thierry Sévenet<sup>§</sup>

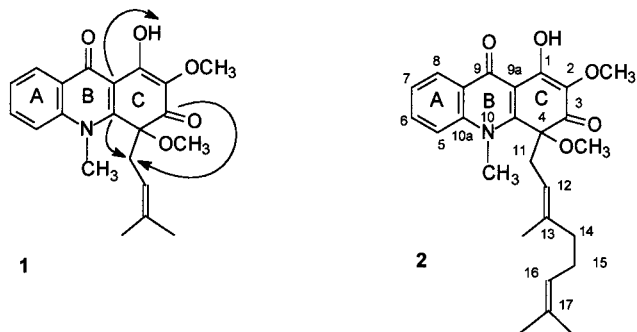
Laboratory of Pharmacognosy, Department of Pharmacy, University of Athens Panepistimiopolis Zografou, GR-15771 Athens, Greece, Laboratoire de Pharmacognosie de l'Université René Descartes, UMR au CNRS No. 8638, Faculté de Pharmacie, 4 Avenue de l'Observatoire, F-75006 Paris, France, and ICSN du CNRS, F-91190 Gif-sur-Yvette, France

Received August 18, 1999

Two new alkaloids, megistophylline I (**1**) and megistophylline II (**2**), were isolated from the bark of *Sarcomelicope megistophylla*. Their structures have been elucidated on the basis of MS and NMR data.

*Sarcomelicope megistophylla* Hartley (Rutaceae) is a small to medium-sized tree, 8–12 m high, easily recognized by its large (up to 34 cm long) pubescent leaves. Hartley<sup>1</sup> described it as endemic to the region of Néaoua, New Caledonia. Recently we described chemical constituents of its leaves.<sup>2,3</sup> In a continuation of our studies of the genus *Sarcomelicope*,<sup>4</sup> we report here a phytochemical study of the bark of *S. megistophylla*. Two new acridone alkaloids, megistophylline I (**1**) and megistophylline II (**2**), were isolated from the ether and dichloromethane bark extracts. The structure of the two novel alkaloids was deduced from <sup>1</sup>H and <sup>13</sup>C NMR spectral data and by interpretation of COSY 45°, COSY–LR, DEPT 135°, HMQC, HMQC–TOCSY, and HMBC experiments.

Megistophylline I (**1**) was obtained as a colorless amorphous compound, and its empirical formula was determined by HRMS as C<sub>21</sub>H<sub>23</sub>NO<sub>5</sub>. The UV spectrum was suggestive of an acridone derivative. The <sup>1</sup>H NMR spectrum indicated four aromatic protons associated with a nonsubstituted ring A in an acridone-derived skeleton, one 1,1-dimethylallyl side chain, two OMe groups, one NMe group, and one hydrogen-bonded phenolic OH group. The <sup>13</sup>C NMR confirmed the above observations and showed the presence of two carbonyl groups. One (δ 178.1) is included in the quinolone skeleton, and the second corresponds to a conjugated ketone (δ 186). The <sup>1</sup>H and <sup>13</sup>C NMR values observed were closely related to those previously reported for a compound obtained by m-CPBA oxidation of glyco-citrine.<sup>5</sup> The only significant differences were the absence of the proton at C-2 and the presence of a methoxy group. These data were consistent with significant chemical shift differences observed for the signals of C-1, C-2, and C-3. Further information on the structure of **1** was obtained from the long-range C–H correlations in the HMBC spectrum (Figure 1). Three-bond correlations between the two methylene protons at δ 3.18 and 2.93 and the 4-tertiary carbon at δ 155.0, on one hand, and the ketonic carbonyl group at δ 186.0, on the other hand, permitted placement of the 1,1-dimethylallyl side chain at C-4 and the carbonyl group at C-3 of the acridone ring C. Finally, observation of three bond correlations between the strongly deshielded hydrogen-bonded hydroxyl at δ 16.22 and the 9a-quaternary carbon at δ 109.2 placed the phenolic hydroxyl group at C-1. Consequently, structure **1** was attributed to this new natural product for which we propose



**Figure 1.** Selected HMBC correlations for megistophylline I (**1**) and megistophylline II (**2**).

the trivial name megistophylline I. The absolute configuration of the chiral center at C-4 could not be determined, due to the small amount of product isolated.

Megistophylline II (**2**) was obtained as a colorless amorphous compound with the empirical formula C<sub>26</sub>H<sub>31</sub>NO<sub>5</sub>. The UV spectrum was similar to that of **1**. Comparison of the <sup>1</sup>H and <sup>13</sup>C NMR data of **2** with those of **1** indicated that the 1,1-dimethylallyl side chain in **1** was replaced by a geranyl group in **2**. The presence of the geranyl group was further confirmed by the EIMS spectrum, where two prominent fragment ions (*m/z* 368 and 301) were observed. Finally, the HMBC spectrum exhibited the same cross signals as those observed in **1**.

Seven known acridones, melicopicine,<sup>6,7</sup> melicopine,<sup>8</sup> melicopidine,<sup>8,9</sup> normelicopidine,<sup>10,11</sup> normelicopine,<sup>12</sup> arborinine,<sup>13</sup> and 1,2,3-trimethoxy-10-methylacridone;<sup>3,6</sup> three pyranacridones, acronycine,<sup>6,14</sup> 12-demethylacronycine,<sup>12</sup> and noracronycine;<sup>15</sup> two furoquinolines, dictamine<sup>9</sup> and acronycidine;<sup>16</sup> one pyranocoumarin, seselin;<sup>17</sup> and the alkaloid fareanine<sup>18</sup> were also obtained. The two new alkaloids, as well as fareanine,<sup>18</sup> are acridone-derived secondary metabolites characterized by a highly oxygenated C-ring. Compounds with such high oxidation levels belonging to the same series were previously isolated only from *Medicosma fareana*<sup>18</sup> and *Sarcomelicope dogniensis*.<sup>4</sup> Megistophylline II (**2**), is the first example of a C-geranyl acridone. Interestingly, the isolation of 3-geranyloxy-1-hydroxy-4-methoxy-10-methylacridone from *Sarcomelicope leiocarpa*<sup>19</sup> bark was the only previous record of a natural acridone bearing an *O*-geranyl substituent.

### Experimental Section

**General Experimental Procedures.** Optical rotations were measured with a Perkin–Elmer 341 polarimeter. NMR spectra were recorded on Bruker DRX 400 and Bruker AC 200

\* To whom correspondence should be addressed. Tel.: +30-1-7274598. Fax: +30-1-7274594. E-mail: skaltsounis@pharm.uoa.gr.

<sup>†</sup> University of Athens.

<sup>‡</sup> Université René Descartes, Paris.

<sup>§</sup> ICSN du CNRS.

spectrometers [ $^1\text{H}$  (400 and 200 MHz) and  $^{13}\text{C}$  (50 MHz)]; chemical shifts are expressed in parts per million (ppm) downfield to TMS. The 2D NMR experiments were performed using standard Bruker microprograms. EIMS, HP-6890; HRMS, AEI MS-902 spectrometer.

**Plant Material.** Plant material was collected at Néaoua (New Caledonia) in May 1984. Herbarium samples (Pusset-Chauvière 261) are deposited in the herbaria of the Centre ORSTOM of Nouméa, New Caledonia.

**Extraction and Isolation of Alkaloids.** Dried, pulverized bark of *Sarcomelicope megistophylla* (1 kg) was basified with  $\text{NH}_4\text{OH}$  and extracted with ether ( $4 \times 2$  L) and repeated with  $\text{CH}_2\text{Cl}_2$  ( $4 \times 2$  L). The residue of the  $\text{CH}_2\text{Cl}_2$  extract was washed with 1N HCl. The aqueous solutions were basified with  $\text{NH}_4\text{OH}$  to pH 10 and extracted with  $\text{CH}_2\text{Cl}_2$  ( $3 \times 1$  L). After drying over  $\text{Na}_2\text{SO}_4$ , the organic solvent was removed in vacuo to give a crude alkaloid mixture (14 g), which was subjected to vacuum liquid chromatography (VLC) over S gel using an EtOAc-hexane gradient to give six fractions.

Fractions 2–3 were chromatographed (with a cyclohexane–EtOAc gradient, Si gel 0.015–0.04 mm) to afford 1.2 g melicopine<sup>6,7</sup> and 0.8 g acronycine.<sup>6,14</sup> Fraction 4 was chromatographed to afford 1.2 g melicopidine,<sup>8,9</sup> 1.8 g melicopine,<sup>8</sup> 0.7 g normelicopidine,<sup>10,11</sup> 0.6 g normelicopine,<sup>12</sup> and 2.4 g acronycine.<sup>6,14</sup> Fraction 5 was chromatographed to afford 150 mg 1,2,3-trimethoxy-10-methylacridone,<sup>3,6</sup> 20 mg megistophylline I (1), 10 mg megistophylline II (2), and 5 mg fareanine. Using the same procedure as above, the residue of the ether extract afforded 30 mg dictamine,<sup>9</sup> 300 mg melicopine,<sup>6,7</sup> 10 mg noracronycine,<sup>15</sup> 9 mg seseline,<sup>17</sup> 0.13 g melicopidine,<sup>8,9</sup> 13 mg melicopine,<sup>8</sup> 90 mg acronycine,<sup>6,14</sup> 30 mg acronycidine,<sup>16</sup> 30 mg 12-demethylacronycine,<sup>12</sup> 70 mg normelicopidine,<sup>10,11</sup> 50 mg normelicopine,<sup>12</sup> 60 mg arborinine,<sup>13</sup> and 40 mg 1,2,3-trimethoxy-10-methylacridone.<sup>3,6</sup> All the compounds isolated were identified by comparison of their spectral (NMR, MS, and UV) and chromatographic data with those of authentic samples.

**Megistophylline I (1):**  $[\alpha]_{\text{D}}^{25} +32.9^\circ$  (*c* 0.26,  $\text{CHCl}_3$ ); UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 301 (3.95), 342 (3.88), 407 (sh) nm; IR (KBr)  $\nu_{\text{max}}$  3350, 1630, 1580, 1520, 1480  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 MHz)  $\delta$  16.22 (1H, s, HO-1), 3.84 (3H, s,  $\text{CH}_3\text{O}-2$ ), 3.23 (3H, s,  $\text{CH}_3\text{O}-4$ ), 7.85 (1H, d,  $J = 8.3$  Hz, H-5), 7.94 (1H, td,  $J = 8.3$ , 1.5 Hz, H-6), 7.65 (1H, t,  $J = 8.3$  Hz, H-7), 8.57 (1H, dd,  $J = 8.3$ , 1.5 Hz, H-8), 4.52 (3H, s, N- $\text{CH}_3$ ), 2.93 (1H, dd,  $J = 14.5$ , 8.3 Hz, H-11a), 3.18 (1H, dd,  $J = 14.5$ , 8.3 Hz, H-11b), 4.78 (1H, t,  $J = 8.3$  Hz, H-12), 1.38 (3H, s,  $\text{CH}_3-13$ ), 1.64 (3H, s,  $\text{CH}_3-13$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 50 MHz)  $\delta$  164.0 (C-1), 132.3 (C-2), 60.0 ( $\text{CH}_3\text{O}-2$ ), 186.0 (C-3), 85.5 (C-4), 53.5 ( $\text{CH}_3\text{O}-4$ ), 155.0 (C-4a), 116.4 (C-5), 134.4 (C-6), 126.2 (C-7), 126.5 (C-8), 124.7 (C-8a), 178.1 (C-9), 109.2 (C-9a), 141.2 (C-10a), 37.1 (N- $\text{CH}_3$ ), 40.0 (C-11), 114.0 (C-12), 137.8 (C-13), 26.0 ( $\text{CH}_3-13$ ), 17.0 ( $\text{CH}_3-13$ ); HRMS found 369.15762 (calcd 369.15762); EIMS  $m/z$  (rel int)  $[\text{M}]^+$  369 (22), 301 (30), 286 (100), 272 (20).

**Megistophylline II (2):**  $[\alpha]_{\text{D}}^{25} +11.26^\circ$  (*c* 0.10,  $\text{CHCl}_3$ ); UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 301 (4.36), 338 (3.88), 404 (sh) nm; IR (KBr)  $\nu_{\text{max}}$  3350, 1632, 1580, 1520, 1480  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 MHz)  $\delta$  16.20 (1H, s, HO-1), 3.85 (3H, s,  $\text{CH}_3\text{O}-2$ ), 3.23 (3H, s,  $\text{CH}_3\text{O}-4$ ), 7.85 (1H, d,  $J = 8.3$  Hz, H-5), 7.92 (1H, td,  $J = 8.3$ , 1.5 Hz, H-6), 7.65 (1H, t,  $J = 8.3$  Hz, H-7), 8.55 (1H, dd,  $J = 8.3$ , 1.5 Hz, H-8), 4.52 (3H, s, N- $\text{CH}_3$ ), 2.97 (1H, dd,  $J = 14.5$ , 8.3 Hz, H-11a), 3.09 (1H, dd,  $J = 14.5$ , 8.3 Hz, H-11b), 4.81 (1H, t,  $J = 8.3$  Hz, H-12), 1.34 (3H, s,  $\text{CH}_3-13$ ), 1.83 (2H, br s, H-14), 1.83 (2H, br s, H-15), 4.93 (1H, br s, H-16), 1.62 (3H, s,  $\text{CH}_3-17$ ), 1.51 (3H, s,  $\text{CH}_3-17$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 50 MHz)  $\delta$  165.0 (C-1), 132.6 (C-2), 60.0 ( $\text{CH}_3\text{O}-2$ ), 186.2 (C-3), 85.8 (C-4), 54.2 ( $\text{CH}_3\text{O}-4$ ), 155.1 (C-4a), 116.5 (C-5), 134.4 (C-6), 125.9 (C-7), 126.8 (C-8), 124.0 (C-8a), 178.2 (C-9), 108.0 (C-9a), 141.4 (C-10a), 37.3 (N- $\text{CH}_3$ ), 39.4 (C-11), 114.0 (C-12), 141.5 (C-13), 16.0 ( $\text{CH}_3-13$ ), 39.6 (C-14), 26.2 (C-15), 123.3 (C-16), 131.8 (C-17), 25.9 ( $\text{CH}_3-17$ ), 17.3 ( $\text{CH}_3-17$ ); HRMS found 437.2202 (calcd 437.2202); EIMS  $m/z$  (rel int)  $[\text{M}]^+$  437 (12), 368 (10), 301 (30), 286 (100), 272 (20), 258 (20), 242 (24).

## References and Notes

- Hartley, T. G. *Bull. Mus. Natl. Hist. Nat. Sec. B. Adansonia (Bot. Phytochim.)* **1986**, *8*, 183–189.
- Fokialakis, N.; Mitaku, S.; Mikros, E.; Skaltsounis, A. L.; Tillequin, F.; Sévenet, T. *Phytochemistry*, in press.
- Skaltsounis, A. L.; Seddrati, L.; Tillequin, F.; Koch, M.; Pusset, J.; Sévenet, T. *Nat. Prod. Lett.* **1995**, *5*, 281–287.
- Mitaku, S.; Skaltsounis, A. L.; Tillequin, F.; Koch, M.; Pusset, J.; Sévenet, T. *Nat. Prod. Lett.* **1995**, *7*, 219–225.
- Ito, C.; Ono, T.; Hatano, K.; Furukawa, H. *Chem. Pharm. Bull.* **1993**, *41*, 383–385.
- Cougé, B.; Tillequin, F.; Koch, M.; Sévenet, T. *Plant. Méd. Phytothér.* **1980**, *14*, 208–212.
- Mitaku, S.; Pusset, J. *Plant. Méd. Phytothér.* **1988**, *22*, 83–87.
- Tillequin, F.; Baudouin, G.; Koch, M.; Sévenet, T. *J. Nat. Prod.* **1980**, *43*, 498–502.
- Mitaku, S.; Skaltsounis, A. L.; Tillequin, F.; Koch, M.; Pusset, J.; Chauvière, G. *J. Nat. Prod.* **1986**, *49*, 1091–1095.
- Svoboda, G. H. *Lloydia* **1966**, *29*, 206–224.
- Mitaku, S.; Skaltsounis, A. L.; Tillequin, F.; Koch, M.; Pusset, J. *Ann. Pharm. Fr.* **1989**, *47*, 149–156.
- Funayama, S.; Cordell, G. A. *J. Nat. Prod.* **1984**, *47*, 285–291.
- Reisch, J.; Adesina, S. K.; Bergenthal, D. *Pharmazie* **1985**, *40*, 811–812.
- Mitaku, S.; Skaltsounis, A. L.; Tillequin, F.; Koch, M.; Pusset, J.; Chauvière, G. *Heterocycles* **1987**, *26*, 2057–2063.
- Rózsa, Z.; Szendrei, K.; Kovacs, Z.; Novák, I.; Minker, E.; Reisch, J. *Phytochemistry* **1978**, *17*, 169–170.
- Skaltsounis, A. L.; Tillequin, F.; Koch, M. *J. Nat. Prod.* **1983**, *46*, 732–735.
- Murray, R. D. H.; Ballantyne, M. M.; Mathai, K. P. *Tetrahedron* **1971**, *27*, 1247–1251.
- Habtemarian, S.; Waterman, P. G.; Hartley, T. G. *Phytochemistry* **1996**, *43*, 291–294.
- Baudouin, G.; Tillequin, F.; Koch, M.; Dau, E. T. H.; Guilhem, J.; Pusset, J.; Chauvière, G. *J. Nat. Prod.* **1985**, *48*, 260–265.

NP9904096